

UNIVERSIDADE FEDERAL DE ITAJUBÁ – UNIFEI
INSTITUTO DE FÍSICA E QUÍMICA
PROGRAMA DE PÓS-GRADUAÇÃO EM MATERIAIS PARA
ENGENHARIA

Microesferas de Poliglicerol Contendo Fumarato de
Dimetila e Curcumina para Aplicação na Terapia da
Esclerose Múltipla

Itajubá/MG
2019

UNIVERSIDADE FEDERAL DE ITAJUBÁ – UNIFEI
INSTITUTO DE FÍSICA E QUÍMICA
PROGRAMA DE PÓS-GRADUAÇÃO EM MATERIAIS PARA
ENGENHARIA

Priscila Veloso da Silva

Microesferas de Poliglicerol Contendo Fumarato de
Dimetila e Curcumina para Aplicação na Terapia da
Esclerose Múltipla

Defesa de Tese submetida ao Programa de Pós-Graduação em Materiais para Engenharia como requisito para a obtenção do título de Doutor em Ciências em Engenharia de Materiais.

Área de Concentração: Não-Metals

Orientador: Prof. Dr. Alvaro Antonio Alencar de Queiroz

Itajubá/MG

2019

Resumo

A esclerose múltipla é uma doença autoimune e crônica caracterizada pela desmielinização dos neurônios no sistema nervoso central. Trata-se de uma doença incapacitante ao longo do tempo e cujas terapias podem provocar diversos efeitos colaterais. Este trabalho propõe uma alternativa ao tratamento da esclerose múltipla a partir do desenvolvimento de microesferas de dendrímero de poliglicerol carreadoras de fumarato de dimetila e de curcumina. Inicialmente, as microesferas foram caracterizadas por meio de MEV, FTIR, TGA e DSC. Foram determinados o grau de intumescimento e a energia de ativação das microesferas de PGLD (G10, G25, G50, G100 e G200). Os resultados dessas análises mostraram que as microesferas porosas, G100 e G200, de menor e maior tamanho, respectivamente, eram as mais adequadas para a formação dos compostos. Foram avaliadas a porosimetria desses grupos e a biocompatibilidade por meio de ensaios de citotoxicidade e hemocompatibilidade *in vitro*. O estudo teórico de ancoragem molecular mostrou que a formação dos compostos foi espontânea, sendo que, o composto com curcumina (-23,8 kJ/mol) apresentou maior afinidade em comparação ao composto com fumarato de dimetila (-11,3 kJ/mol). Os compostos, DMF-PGLD e CUR-PGLD, foram preparados e, posteriormente, caracterizados por FTIR e avaliados os perfis de liberação. Os espectros de FTIR sugeriram a interação dos compostos devido ao aparecimento de bandas de absorção características das moléculas precursoras nos compostos. Em geral, o estudo de liberação determinou o mecanismo de transporte super caso II. Foi observada uma retenção maior nos compostos com curcumina, isso porque as curvas se apresentaram menos acentuadas ou com liberação mais tardia. Em conclusão, sugere-se que a formação de compostos entre microesferas porosas de dendrímero de poliglicerol (G100 e G200) e os ativos (fumarato de dimetila e curcumina) seja uma alternativa para a liberação controlada no tratamento da esclerose múltipla, com o intuito de diminuir os efeitos colaterais. Entretanto, os compostos com curcumina tiveram resultados mais favoráveis a sua aplicação, em virtude de sua maior afinidade e retenção.

Palavras-chave: Esclerose múltipla, sistemas de liberação controlada, dendrímero de poliglicerol, fumarato de dimetila, curcumina.

Abstract

Multiple sclerosis is autoimmune and chronic disease characterized for neurons demyelinating and formation of sclerotic lesions within central nervous system. It is a disabling disease over time and whose therapies can cause several side effects. This work proposes an alternative to the treatment of multiple sclerosis from the development of carrier system using microspheres of polyglycerol dendrimer with dimethyl fumarate and curcumin. Initially, the microspheres were characterized by SEM, FTIR, TGA and DSC. The swelling degree and the activation energy of the polyglycerol dendrimer microspheres (G10, G25, G50, G100 and G200) were determined. The results of these analyzes showed that the smaller and larger porous microspheres G100 and G200, respectively, were the most suitable for the formation of the compounds. It was evaluated porosimetry of these groups and biocompatibility by *in vitro* cytotoxicity and hemocompatibility assays. The theoretical study of molecular anchoring showed that the formation of the compounds was spontaneous, the compound with curcumin (-23.8 kJ/mol) had higher affinity compared to the compound with dimethyl fumarate (-11.3 kJ/mol). The compounds, DMF-PGLD and CUR-PGLD, were prepared and subsequently characterized by FTIR and evaluated the release profiles. FTIR spectra suggested the interaction of the compounds due to the appearance of absorption bands characteristic of the precursor molecules in the compounds. The release study determined the super case II transport mechanism. A higher retention was observed in the compounds with curcumin, because the curves were less pronounced or with later release. In conclusion, it is suggested that the formation of compounds between polyglycerol dendrimer (G100 and G200) porous microspheres and active (dimethyl fumarate and curcumin) are an alternative for controlled release in the treatment of multiple sclerosis. However the compounds with curcumin had more favorable results to their application, due to affinity and retention.

Key-words: Multiple sclerosis, controlled release system, polyglycerol dendrimer, dimethyl fumarate, curcumin.

APÊNDICE A - Produção Científica

- 1) SILVA, P. V.; QUEIROZ, A. A. A. Long term multiple sclerosis drug delivery using dendritic polyglycerol flower-like microspheres. *Journal of Biomaterials Science-Polymer Edition*, v. 1, p. 1-13, 2019.

- 2) QUEIROZ, A. A. A.; DETTORI, L. G.; SILVA, P. V.; RODRIGUES, R. R. Nanoesferas ou microesferas de polímero verde transportadoras de curcumina, curcuminóides e seus derivados. 2016, Brasil. Patente: Privilégio de Inovação. Número do registro: BR1020160275601, Instituição de registro: INPI - Instituto Nacional da Propriedade Industrial. Depósito: 24/11/2016

REFERÊNCIAS

1. Tabansky, I. *et al.* Advancing drug delivery systems for the treatment of multiple sclerosis. *Immunol. Res.* **63**, 58–69 (2015).
2. Calabrese, M. *et al.* Exploring the origins of grey matter damage in multiple sclerosis. *Nat. Rev. Neurosci.* **16**, 147 (2015).
3. Murphy, K. & Walport, M. *Imunobiologia de Janeway TT - Immunobiology Janeway.* 908 (2010).
4. Dendrou, C. A., Fugger, L. & Friese, M. A. Immunopathology of multiple sclerosis. *Nat. Rev. Immunol.* **15**, 545 (2015).
5. Filho, G. B. Bogliolo Patologia. in 962–964 (Guanabara Koogan, 2011).
6. Frota, E. R. C., Mendes, M. F. & Vasconcelos, C. C. F. Recomendações no tratamento da esclerose múltipla e neuromielite óptica. in 236 (Editora e Eventos Omnifarma, 2016).
7. Banks, W. A. From blood–brain barrier to blood–brain interface: new opportunities for CNS drug delivery. *Nat. Rev. Drug Discov.* **15**, 275 (2016).
8. Palanichamy, A. *et al.* Immunoglobulin class-switched B cells provide an active immune axis between CNS and periphery in multiple sclerosis. *Sci. Transl. Med.* **6**, 248ra106–248ra106 (2014).
9. Rahmanzadeh, R., Weber, M. S., Bruck, W., Navardi, S. & Sahraian, M. A. B cells in multiple sclerosis therapy-A comprehensive review. *Acta Neurol. Scand.* **137**, 544–556 (2018).
10. Budnik, V., Ruiz-Cañada, C. & Wendler, F. Extracellular vesicles round off communication in the nervous system. *Nat. Rev. Neurosci.* **17**, 160 (2016).
11. Xie, L., Li, X.-K. & Takahara, S. Curcumin has bright prospects for the treatment of multiple sclerosis. *Int. Immunopharmacol.* **11**, 323–330 (2011).
12. Huang, G., Wang, Y., Vogel, P. & Chi, H. Control of IL-17 receptor signaling and tissue inflammation by the p38 α –MKP-1 signaling axis in a mouse model of multiple sclerosis. *Sci. Signal.* **8**, ra24–ra24 (2015).
13. Cao, Y. *et al.* Functional inflammatory profiles distinguish myelin-reactive T cells from patients with multiple sclerosis. *Sci. Transl. Med.* **7**, 287ra74 LP–287ra74 (2015).
14. Neil, S. *et al.* Oral administration of the nitroxide radical TEMPOL exhibits immunomodulatory and therapeutic properties in multiple sclerosis models. *Brain. Behav. Immun.* **62**, 332–343 (2017).
15. Fugger, L., Friese, M. A. & Bell, J. I. From genes to function: the next challenge to understanding multiple sclerosis. *Nat. Rev. Immunol.* **9**, 408 (2009).
16. World Health Organization. Federation MSI. Atlas of MS 2013. in 28 (2013).
17. Leray, E., Moreau, T., Fromont, A. & Edan, G. Epidemiology of multiple sclerosis.

- Rev. Neurol. (Paris)*. **172**, 3–13 (2016).
18. Vasconcelos, C. C. F., Thuler, L. C. S., Rodrigues, B. C., Calmon, A. B. & Alvarenga, R. M. P. Multiple sclerosis in Brazil: A systematic review. *Clin. Neurol. Neurosurg.* **151**, 24–30 (2016).
 19. da Gama Pereira, A. B. C. N., Sampaio Lacativa, M. C., da Costa Pereira, F. F. C. & Papais Alvarenga, R. M. Prevalence of multiple sclerosis in Brazil: A systematic review. *Mult. Scler. Relat. Disord.* **4**, 572–579 (2015).
 20. Finkelsztejn, A., Lopes, J. S., Noal, J. & Finkelsztejn, J. M. The prevalence of multiple sclerosis in Santa Maria, Rio Grande do Sul, Brazil. *Arq. Neuropsiquiatr.* **72**, 104–106 (2014).
 21. Chitnis, T., Glanz, B., Jaffin, S. & Healy, B. Demographics of pediatric-onset multiple sclerosis in an MS center population from the Northeastern United States. *Mult. Scler.* **15**, 627–631 (2009).
 22. Krupp, L. B., Banwell, B. & Tenenbaum, S. Consensus definitions proposed for pediatric multiple sclerosis and related disorders. *Neurology* **68**, S7-12 (2007).
 23. McCombe, P. A. & Stenager, E. Female infertility and multiple sclerosis: is this an issue? *Multiple sclerosis (Houndmills, Basingstoke, England)* **21**, 5–7 (2015).
 24. Organização Mundial da Saúde. Available at: <http://www.who.int/classifications/icd/en/>. (Acessado: 12º abril 2018)
 25. Ministério da Saúde. Available at: http://www.datasus.gov.br/cid10/V2008/WebHelp/g00_g99.htm. (Acessado: 13º abril 2018)
 26. Ministério da Saúde. Available at: <http://tabnet.datasus.gov.br/cgi/tabcgi.exe?sim/cnv/mat10uf.def>. (Acessado: 21º setembro 2018)
 27. Secretaria de Previdência - Ministério da Fazenda. Available at: <http://www.previdencia.gov.br/dados-abertos/dados-abertos-previdencia-social/>. (Acessado: 14º setembro 2018)
 28. ANZgene, M. S. G. C. Genome-wide association study identifies new multiple sclerosis susceptibility loci on chromosomes 12 and 20. *Nat. Genet.* **41**, 824–828 (2009).
 29. Tullman, M. J. A review of current and emerging therapeutic strategies in multiple sclerosis. *Am. J. Manag. Care* **19**, S21-7 (2013).
 30. Katz Sand, I. Classification, diagnosis, and differential diagnosis of multiple sclerosis. *Curr. Opin. Neurol.* **28**, 193–205 (2015).
 31. Winkelmann, A., Loebermann, M., Reisinger, E. C., Hartung, H.-P. & Zettl, U. K. Disease-modifying therapies and infectious risks in multiple sclerosis. *Nat. Rev. Neurol.* **12**, 217–233 (2016).
 32. Brownlee, W. J., Hardy, T. A., Fazekas, F. & Miller, D. H. Diagnosis of multiple

- sclerosis: progress and challenges. *Lancet (London, England)* **389**, 1336–1346 (2017).
33. Polman, C. H. *et al.* Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann. Neurol.* **69**, 292–302 (2011).
 34. Ebers, G. C., Heigenhauser, L., Daumer, M., Lederer, C. & Noseworthy, J. H. Disability as an outcome in MS clinical trials. *Neurology* **71**, 624–631 (2008).
 35. Miller, D. H. & Leary, S. M. Primary-progressive multiple sclerosis. *Lancet. Neurol.* **6**, 903–912 (2007).
 36. Kira, J. Neuromyelitis optica and asian phenotype of multiple sclerosis. *Ann. N. Y. Acad. Sci.* **1142**, 58–71 (2008).
 37. Gafson, A., Giovannoni, G. & Hawkes, C. H. The diagnostic criteria for multiple sclerosis: From Charcot to McDonald. *Mult. Scler. Relat. Disord.* **1**, 9–14 (2012).
 38. McDonald, W. I. *et al.* Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. *Ann. Neurol.* **50**, 121–127 (2001).
 39. Polman, C. H. *et al.* Diagnostic criteria for multiple sclerosis: 2005 revisions to the “McDonald Criteria”. *Ann. Neurol.* **58**, 840–846 (2005).
 40. Hawkes, C. H. & Giovannoni, G. The McDonald Criteria for Multiple Sclerosis: time for clarification. *Mult. Scler.* **16**, 566–575 (2010).
 41. MINISTÉRIO DA, SAÚDE, S. D. A. À. & SECRETARIA DE CIÊNCIA, T. E. I. E. *Portaria Conjunta nº 10, de 02 de abril de 2018.* 27 (2018).
 42. Gabr, R. E., Tefera, G. B., Allen, W. J., Pednekar, A. S. & Narayana, P. A. GRAPE: a graphical pipeline environment for image analysis in adaptive magnetic resonance imaging. *Int. J. Comput. Assist. Radiol. Surg.* **12**, 449–457 (2017).
 43. Haaga, John R.; Dogra, Vikram S.; Forsting, Michael; Gikeson, Robert C.; Ha, Hyun Kwon; Sundaram, M. *TC e RM - Uma abordagem do corpo humano completo.* (2011).
 44. Westbrook, C., Roth, C. K. & Talbot, J. Ressonância magnética: aplicações práticas. in (org. Fernandes, M. V. S. [Tradução]) 373 (Guanabara Koogan, 2016).
 45. Moore, F. G. A. & Levental, M. The Usefulness of Gadolinium-Enhanced Images on a Follow-up Magnetic Resonance Image in Suspected Multiple Sclerosis. *Can. Assoc. Radiol. J.* **64**, 358–362 (2013).
 46. Mejia, A. F. *et al.* Statistical estimation of T1 relaxation times using conventional magnetic resonance imaging. *Neuroimage* **133**, 176–188 (2016).
 47. Santos, M. A. R. *et al.* Contribuição do Mismatch Negativity na avaliação cognitiva de indivíduos portadores de esclerose múltipla. *Rev. Bras. Otorrinolaringol.* **72**, 800–807 (2006).
 48. Leocani, L. & Comi, G. Clinical neurophysiology of multiple sclerosis. *Handb. Clin. Neurol.* **122**, 671–679 (2014).
 49. Domingues, R. B. *et al.* The cerebrospinal fluid in multiple sclerosis: far beyond the

- bands. *Einstein (São Paulo)* **15**, 100–104 (2017).
50. Paty, D. W. *et al.* Management of relapsing-remitting multiple sclerosis: diagnosis and treatment guidelines. *Eur. J. Neurol.* **6**, S1–S35 (1999).
 51. Soelberg Sorensen, P. Safety concerns and risk management of multiple sclerosis therapies. *Acta Neurol. Scand.* **136**, 168–186 (2017).
 52. Pozzilli, C. & Pugliatti, M. An overview of pregnancy-related issues in patients with multiple sclerosis. *Eur. J. Neurol.* **22 Suppl 2**, 34–39 (2015).
 53. Deshmukh, V. A. *et al.* A regenerative approach to the treatment of multiple sclerosis. *Nature* **502**, 327–332 (2013).
 54. Di Filippo, M. *et al.* Persistent activation of microglia and NADPH oxidase drive hippocampal dysfunction in experimental multiple sclerosis. *Sci. Rep.* **6**, 20926 (2016).
 55. Gholamzad, M. *et al.* A comprehensive review on the treatment approaches of multiple sclerosis: currently and in the future. *Inflamm. Res.* (2018). doi:10.1007/s00011-018-1185-0
 56. Etemadifar, M., Janghorbani, M. & Shaygannejad, V. Comparison of Betaferon, Avonex, and Rebif in treatment of relapsing-remitting multiple sclerosis. *Acta Neurol. Scand.* **113**, 283–287 (2006).
 57. Comissão Nacional de Incorporação de Tecnologias no SUS, C. *Betainterferonas no tratamento da esclerose múltipla remitente-recorrente (EMRR)*. 1–50 (2016).
 58. Furber, K. L. *et al.* Advances in the treatment of relapsing–remitting multiple sclerosis: the role of pegylated interferon β -1a. *Degener. Neurol. Neuromuscul. Dis.* **7**, 47–60 (2017).
 59. ANVISA. Available at: http://www.anvisa.gov.br/datavisa/fila_bula/frmVisualizarBula.asp?pNuTransacao=24329322016&pIdAnexo=3980838. (Acessado: 6º maio 2018)
 60. Comissão Nacional de Incorporação de Tecnologias no SUS, C. *Natalizumabe 300mg (Tysabri®) para Esclerose Múltipla Remitente Recorrente em Segunda Linha de Trabalho*. 1–32 (2013).
 61. Pardo, G. & Jones, D. E. The sequence of disease-modifying therapies in relapsing multiple sclerosis: safety and immunologic considerations. *J. Neurol.* **264**, 2351–2374 (2017).
 62. Olsson, T. *et al.* Anti-JC virus antibody prevalence in a multinational multiple sclerosis cohort. *Mult. Scler.* **19**, 1533–1538 (2013).
 63. Tuohy, O. *et al.* Alemtuzumab treatment of multiple sclerosis: long-term safety and efficacy. *J. Neurol. Neurosurg. Psychiatry* **86**, 208–215 (2015).
 64. Comissão Nacional de Incorporação de Tecnologias no SUS, C. *Alemtuzumabe no tratamento da esclerose múltipla remitente recorrente após falha terapêutica com betainterferona ou glatirâmer*. 1–32 (2017).
 65. Kim, A. P. & Baker, D. E. Daclizumab. *Hosp. Pharm.* **51**, 928–939 (2016).

66. Comissão Nacional de Incorporação de Tecnologias no SUS, C. *Teriflunomida para primeira linha de tratamento da esclerose múltipla remitente recorrente*. 1–45 (2017).
67. Comissão Nacional de Incorporação de Tecnologias no SUS, C. *Fumarato de dimetila no tratamento da esclerose múltipla remitente recorrente após falha com betainterferona ou glatirâmer*. 1–99 (2017).
68. Jattinagoudar, L., Meti, M., Nandibewoor, S. & Chimatadar, S. Evaluation of the binding interaction between bovine serum albumin and dimethyl fumarate, an anti-inflammatory drug by multispectroscopic methods. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **156**, 164–171 (2016).
69. National Multiple Sclerosis Society, N. *Vitamins, Minerals & Herbs in MS An Introduction*. (2015).
70. Sintzel, M. B., Rametta, M. & Reder, A. T. Vitamin D and Multiple Sclerosis: A Comprehensive Review. *Neurol. Ther.* **7**, 59–85 (2018).
71. Becker, J. *et al.* Hypovitaminosis D association with disease activity in relapsing remitting multiple sclerosis in Brazil. *J. Neurol. Sci.* **363**, 236–239 (2016).
72. McLaughlin, L. *et al.* Vitamin D for the treatment of multiple sclerosis: a meta-analysis. *J. Neurol.* (2018). doi:10.1007/s00415-018-9074-6
73. Schmitz, K. *et al.* “Disease modifying nutricals” for multiple sclerosis. *Pharmacol. Ther.* **148**, 85–113 (2015).
74. Handunnetthi, L., Ramagopalan, S. V & Ebers, G. C. Multiple sclerosis, vitamin D, and HLA-DRB1*15. *Neurology* **74**, 1905–1910 (2010).
75. Ammon, H. P. & Wahl, M. A. Pharmacology of Curcuma longa. *Planta Med.* **57**, 1–7 (1991).
76. Sawant, V. J., Bamane, S. R., Shejwal, R. V & Patil, S. B. Comparison of drug delivery potentials of surface functionalized cobalt and zinc ferrite nanohybrids for curcumin in to MCF-7 breast cancer cells. *J. Magn. Magn. Mater.* **417**, 222–229 (2016).
77. Agrawal, D. K. & Mishra, P. K. Curcumin and its analogues: potential anticancer agents. *Med. Res. Rev.* **30**, 818–860 (2010).
78. Nayunigari, M. K., Maity, A., Agarwal, S. & Gupta, V. K. Curcumin–malic acid based green copolymers for control of scale and microbiological growth applications in industrial cooling water treatment. *J. Mol. Liq.* **214**, 400–410 (2016).
79. Panahi, Y. *et al.* Evidence of curcumin and curcumin analogue effects in skin diseases: A narrative review. *J. Cell. Physiol.* **0**, (2018).
80. Seyedzadeh, M. H. *et al.* Study of curcumin immunomodulatory effects on reactive astrocyte cell function. *Int. Immunopharmacol.* **22**, 230–235 (2014).
81. Tegenge, M. A. *et al.* Curcumin protects axons from degeneration in the setting of local neuroinflammation. *Exp. Neurol.* **253**, 102–110 (2014).
82. Potter, K. A. *et al.* Curcumin-releasing mechanically adaptive intracortical implants improve the proximal neuronal density and blood-brain barrier stability. *Acta Biomater.*

- 10**, 2209–2222 (2014).
83. Anand, P., Kunnumakkara, A. B., Newman, R. A. & Aggarwal, B. B. Bioavailability of curcumin: problems and promises. *Mol. Pharm.* **4**, 807–818 (2007).
 84. Xie, L. *et al.* Amelioration of experimental autoimmune encephalomyelitis by curcumin treatment through inhibition of IL-17 production. *Int. Immunopharmacol.* **9**, 575–581 (2009).
 85. Ghalandarlaki, N., Alizadeh, A. M. & Ashkani-Esfahani, S. Nanotechnology-applied curcumin for different diseases therapy. *Biomed Res. Int.* **2014**, 394264 (2014).
 86. Baj, T. & Seth, R. Role of Curcumin in Regulation of TNF-alpha Mediated Brain Inflammatory Responses. *Recent Pat. Inflamm. Allergy Drug Discov.* **12**, 69–77 (2018).
 87. Wang, Y., Wang, C., Zhao, J., Ding, Y. & Li, L. A cost-effective method to prepare curcumin nanosuspensions with enhanced oral bioavailability. *J. Colloid Interface Sci.* **485**, 91–98 (2017).
 88. Anand, P. *et al.* Design of curcumin-loaded PLGA nanoparticles formulation with enhanced cellular uptake, and increased bioactivity in vitro and superior bioavailability in vivo. *Biochem. Pharmacol.* **79**, 330–338 (2010).
 89. Shi, W. *et al.* Synthesis of monofunctional curcumin derivatives, clicked curcumin dimer, and a PAMAM dendrimer curcumin conjugate for therapeutic applications. *Org. Lett.* **9**, 5461–5464 (2007).
 90. Ferreira-Da-Silva, A. L. *et al.* Diretriz para análises de impacto orçamentário de tecnologias em saúde no Brasil. *Cad. Saúde Públicas* **28**, 1223–1238 (2012).
 91. Hartung, D. M., Bourdette, D. N., Ahmed, S. M. & Whitham, R. H. The cost of multiple sclerosis drugs in the US and the pharmaceutical industry: Too big to fail? *Neurology* **84**, 2185–2192 (2015).
 92. Pezzini, B. R., Silva, M. A. S. & Ferraz, H. G. Formas farmacêuticas sólidas orais de liberação prolongada: sistemas monolíticos e multiparticulados. *Rev. Bras. Ciências Farm.* **43**, 491–502 (2007).
 93. Kim, S., Kim, J.-H., Jeon, O., Kwon, I. C. & Park, K. Engineered polymers for advanced drug delivery. *Eur. J. Pharm. Biopharm.* **71**, 420–430 (2009).
 94. Hoffman, A. S. The origins and evolution of “controlled” drug delivery systems. *J. Control. Release* **132**, 153–163 (2008).
 95. Prajapati, S. *et al.* Dendrimers in Drug Delivery, Diagnosis and Therapy: Basics and Potential Applications. *J. Drug Deliv. Ther.* **6**, 67–92 (2016).
 96. Felice, B., Prabhakaran, M. P., Rodriguez, A. P. & Ramakrishna, S. Drug delivery vehicles on a nano-engineering perspective. *Mater. Sci. Eng. C. Mater. Biol. Appl.* **41**, 178–195 (2014).
 97. Hutson, P. H., Clark, J. A. & Cross, A. J. CNS Target Identification and Validation: Avoiding the Valley of Death or Naive Optimism? *Annu. Rev. Pharmacol. Toxicol.* **57**,

- 171–187 (2017).
98. Mitragotri, S., Burke, P. A. & Langer, R. Overcoming the challenges in administering biopharmaceuticals: formulation and delivery strategies. *Nat. Rev. Drug Discov.* **13**, 655–672 (2014).
 99. Kumari, A., Yadav, S. K. & Yadav, S. C. Biodegradable polymeric nanoparticles based drug delivery systems. *Colloids Surf. B. Biointerfaces* **75**, 1–18 (2010).
 100. Barenholz, Y. Doxil®--the first FDA-approved nano-drug: lessons learned. *J. Control. Release* **160**, 117–134 (2012).
 101. Senapati, S., Mahanta, A. K., Kumar, S. & Maiti, P. Controlled drug delivery vehicles for cancer treatment and their performance. *Signal Transduct. Target. Ther.* **3**, 7 (2018).
 102. Park, S.-C. *et al.* Targeting and synergistic action of an antifungal peptide in an antibiotic drug-delivery system. *J. Control. Release* **256**, 46–55 (2017).
 103. Gong, J., Chen, M., Zheng, Y., Wang, S. & Wang, Y. Polymeric micelles drug delivery system in oncology. *J. Control. Release* **159**, 312–323 (2012).
 104. Siddiqi, K. S., Husen, A., Sohrab, S. S. & Yassin, M. O. Recent Status of Nanomaterial Fabrication and Their Potential Applications in Neurological Disease Management. *Nanoscale Res. Lett.* **13**, (2018).
 105. Lu, C.-T. *et al.* Current approaches to enhance CNS delivery of drugs across the brain barriers. *Int. J. Nanomedicine* **9**, 2241–2257 (2014).
 106. Yang, S. K. & Zimmerman, S. C. Water-Soluble Polyglycerol Dendrimers with Two Orthogonally Reactive Core Functional Groups for One-Pot Functionalization. *Macromolecules* **48**, 2504–2508 (2015).
 107. Dervedde, J. *et al.* Dendritic polyglycerol sulfates as multivalent inhibitors of inflammation. *Proc. Natl. Acad. Sci. U. S. A.* **107**, 19679–19684 (2010).
 108. Fernandes, E. G. R. & De Queiroz, A. A. A. A bioconjugated polyglycerol dendrimer with glucose sensing properties. *J. Mater. Sci. Mater. Med.* **20**, 473–479 (2009).
 109. Higa, O., Faria, H. & De Queiroz, A. Polyglycerol dendrimers immobilized on radiation grafted poly-HEMA hydrogels: Surface chemistry characterization and cell adhesion. *Radiat. Phys. Chem.* **98**, 118–123 (2014).
 110. Alonso, M. J. Nanomedicines for overcoming biological barriers. *Biomed. Pharmacother.* **58**, 168–172 (2004).
 111. Uppal, S., Italiya, K. S., Chitkara, D. & Mittal, A. Nanoparticulate-based drug delivery systems for small molecule anti-diabetic drugs: An emerging paradigm for effective therapy. *Acta Biomater.* (2018).
 112. Zhang, X. *et al.* Monitoring local delivery of vancomycin from gelatin nanospheres in zebrafish larvae. *Int. J. Nanomedicine* **13**, 5377–5394 (2018).
 113. Shukla, T. *et al.* Biomedical applications of microemulsion through dermal and transdermal route. *Biomed. Pharmacother.* **108**, 1477–1494 (2018).

114. Zhao, Y.-Z. *et al.* Selection of high efficient transdermal lipid vesicle for curcumin skin delivery. *Int. J. Pharm.* **454**, 302–309 (2013).
115. Jalalvandi, E. & Shavandi, A. In situ-forming and pH-responsive hydrogel based on chitosan for vaginal delivery of therapeutic agents. *J. Mater. Sci. Mater. Med.* **29**, 158 (2018).
116. Tayeb, H. H. & Sainsbury, F. Nanoemulsions in drug delivery: formulation to medical application. *Nanomedicine* **13**, 2507–2525 (2018).
117. El-Zaafarany, G. M. *et al.* A Tailored Thermosensitive PLGA-PEG-PLGA/Emulsomes Composite for Enhanced Oxcarbazepine Brain Delivery via the Nasal Route. *Pharmaceutics* **10**, (2018).
118. Ali, J., Fazil, M., Qumbar, M., Khan, N. & Ali, A. Colloidal drug delivery system: amplify the ocular delivery. *Drug Deliv.* **23**, 710–726 (2016).
119. Del Amo, E. M. *et al.* Pharmacokinetic aspects of retinal drug delivery. *Prog. Retin. Eye Res.* **57**, 134–185 (2017).
120. Emami, S., Siah-Shadbad, M., Adibkia, K. & Barzegar-Jalali, M. Recent advances in improving oral drug bioavailability by cocrystals. *Bioimpacts* **8**, 305–320 (2018).
121. Parthipan, A. K. *et al.* One-step fabrication of bicompartamental microparticles as a dual drug delivery system for Parkinson's disease management. *J. Mater. Sci.* **54**, 730–744 (2019).
122. Freire, M. C. L. C. *et al.* Understanding Drug Release Data through Thermodynamic Analysis. *Mater. (Basel, Switzerland)* **10**, (2017).
123. Costa, P. J. C. da. Avaliação in vitro da bioequivalência de formulações farmacêuticas. *Rev. Bras. Ciências Farm.* **38**, 141–153 (2002).
124. Lopes, C., Lobo, J. & Costa, P. Formas farmacêuticas de liberação modificada: polímeros hidrofílicos. *Rev. Bras. Ciências Farm. - RBCF* **41**, (2005).
125. Dash, S., Murthy, P. N., Nath, L. & Chowdhury, P. Kinetic modeling on drug release from controlled drug delivery systems. *Acta Pol. Pharm.* **67**, 217–223 (2010).
126. Ritger, P. L. & Peppas, N. A. A simple equation for description of solute release II. Fickian and anomalous release from swellable devices. *J. Control. Release* **5**, 37–42 (1987).
127. Korsmeyer, R. W., Gurny, R., Doelker, E., Buri, P. & Peppas, N. A. Mechanisms of solute release from porous hydrophilic polymers. *Int. J. Pharm.* **15**, 25–35 (1983).
128. Oliveira, R. M. P. Microesferas de Ivermectina baseados no Poliglicerol arborescente para o Tratamento da Rosacea. (Universidade Federal de Itajubá, 2018).
129. Brunauer, S., Emmett, P. H. & Teller, E. Adsorption of Gases in Multimolecular Layers. *J. Am. Chem. Soc.* **60**, 309–319 (1938).
130. Barrett, E. P., Joyner, L. G. & Halenda, P. P. The Determination of Pore Volume and Area Distributions in Porous Substances. I. Computations from Nitrogen Isotherms. *J. Am. Chem. Soc.* **73**, 373–380 (1951).

131. ASTM. ASTM F 756-00: in *Standard Practices for Assessment of Haemolytic Properties of Materials*, American Society for Testing and Materials (2000).
132. Dobrovolskaia, M. A. & Neun, B. W. Nanotechnology Characterization Laboratory. in *Analysis of haemolytic properties of nanoparticles: NCL Method ITA-1 (version 1.0)* (National Cancer Institute, 2005).
133. Trott, O. & Olson, A. J. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J. Comput. Chem.* **31**, 455–461 (2010).
134. Morris, G. M. *et al.* AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* **30**, 2785–2791 (2009).
135. Dewar, M. J. S., Zoebisch, E. G., Healy, E. F. & Stewart, J. J. P. Development and use of quantum mechanical molecular models. 76. AM1: a new general purpose quantum mechanical molecular model. [Erratum to document cited in CA103(2):11627f]. *J. Am. Chem. Soc.* **115**, 5348 (1993).
136. Parize, A. *et al.* Impregnation of Chitosan Microspheres with the Natural Dye Curcuma. *Lat. Am. J. Pharm.* **28**, 19–26 (2009).
137. Annane, D. Body temperature in sepsis: a hot topic. *Lancet. Respir. Med.* **6**, 162–163 (2018).
138. Kunwar, A., Barik, A., Priyadarsini, I. & Pandey, R. Absorption and fluorescence studies of curcumin bound to liposome and living cells. **285**, (2007).
139. Klose, D., Siepmann, F., Elkharraz, K., Krenzlin, S. & Siepmann, J. How porosity and size affect the drug release mechanisms from PLGA-based microparticles. *Int. J. Pharm.* **314**, 198–206 (2006).
140. Khalil, B., Hussein, A. & Abud, H. Effect of Crosslinking Agent Ratio and Temperature on Degree of Swelling in Polymer Hydrogels. **52**, (2017).
141. Barbosa, L. C. de A. *Espectroscopia no infravermelho na caracterização de compostos orgânicos*. (2007).
142. Moura, R. M. Avaliação do potencial antiproliferativo do dendrímero de poliglicerol associado ao celecoxibe em linhagens celulares de carcinoma epidermoide de cabeça e pescoço. (Universidade de São Paulo, 2014).
143. Oliveira, M. A., Yoshida, M. I. & Gomes, E. C. de L. Análise Térmica Aplicada a Fármacos e Formulações Farmacêuticas na Indústria Farmacêutica. *Quim. Nov.* **34**, 1224–1230 (2011).
144. Mothé, C. G.; Azevedo, A. D. *Análise Térmica de Materiais*. (2009).
145. Zia, F. *et al.* Synthesis and characterization of chitosan/curcumin blends based polyurethanes. *Int. J. Biol. Macromol.* **92**, 1074–1081 (2016).
146. Schubert, C. *et al.* Can Hyperbranched Polymers Entangle? Effect of Hydrogen Bonding on Entanglement Transition and Thermorheological Properties of Hyperbranched Polyglycerol Melts. *Macromolecules* **49**, 8722–8737 (2016).

147. Gardella, L., Forouharshad, M., Pastorino, L. & Monticelli, O. Hyperbranched PDLA-polyglycerol: A novel additive for tuning PLLA electrospun fiber degradation and properties. *Eur. Polym. J.* **91**, 21–30 (2017).
148. Chen, Z. *et al.* Thermal degradation kinetics study of curcumin with nonlinear methods. *Food Chem.* **155**, 81–86 (2014).
149. Díaz-Celorio, E., Franco, L., Márquez, Y., Rodríguez-Galán, A. & Puiggali, J. Thermal degradation studies on homopolymers and copolymers based on trimethylene carbonate and glycolide units. *Thermochim. Acta* **528**, 23–31 (2012).
150. Felix, F. S., Cides da Silva, L. C., Angnes, L. & Matos, J. R. Thermal behavior study and decomposition kinetics of salbutamol under isothermal and non-isothermal conditions. *J. Therm. Anal. Calorim.* **95**, 877–880 (2009).
151. HT, K. & SC, O. Kinetics of Thermal Degradation of Waste Polypropylene and High-Density Polyethylene. *J. Ind. Eng. Chem.* **11**, 648–656 (2005).
152. Silva, G., Nakamura, N. M., Iha, K. Estudo cinético da decomposição térmica do pentaeritrol-tetranitrado (PETN). *Quim. Nova* **31**, 2060–2064 (2008).
153. Lenchenkov, N. S. *et al.* Characterisation of the size and swelling kinetics of copolymer nano-spheres extracted from an emulsion. *Colloids Surfaces A Physicochem. Eng. Asp.* **535**, 265–273 (2017).
154. Crispim, E. G., , J. F. Piai, A. R. Fajardo, E. R. F. R., , T. U. Nakamura, C. V. N. & A. F. Rubira, E. C. M. Hydrogels based on chemically modified poly(vinyl alcohol) (PVA-GMA) and PVA-GMA/chondroitin sulfate: Preparation and characterization. *Polym. Lett.* **6**, 383–395 (2012).
155. Fajardo, A. R. *et al.* Hydrogel based on an alginate–Ca²⁺/chondroitin sulfate matrix as a potential colon-specific drug delivery system. *RSC Adv.* **2**, 11095–11103 (2012).
156. Yang, Q. Z., Fan, C. J., Yang, X. G., Liao, L. Q. & Liu, L. J. Facile synthesis of biocompatible polyglycerol hydrogel based on epichlorohydrin. *J. Appl. Polym. Sci.* **133**, (2016).
157. Cho, I. S. & Ooya, T. A Supramolecular Hydrogel Based on Polyglycerol Dendrimer-Specific Amino Group Recognition. *Chem. Asian J.* **13**, 1688–1691 (2018).
158. Ninawe, P. R. & Parulekar, S. J. Drug Delivery Using Stimuli-Responsive Polymer Gel Spheres. *Ind. Eng. Chem. Res.* **51**, 1741–1755 (2012).
159. Brazel, C. S. & Peppas, N. A. Modeling of drug release from swellable polymers. *Eur. J. Pharm. Biopharm.* **49**, 47–58 (2000).
160. Aouada, F. A., Muniz, E. C., Vaz, C. M. P. & Mattoso, L. H. C. Correlação entre parâmetros da cinética de intumescimento com características estruturais e hidrofílicas de hidrogéis de poliacrilamida e metilcelulose. *Quim. Nova* **32**, 1482–1490 (2009).
161. Babaye Khorasani, F., Poling-Skutvik, R., Krishnamoorti, R. & Conrad, J. C. Mobility of Nanoparticles in Semidilute Polyelectrolyte Solutions. *Macromolecules* **47**, 5328–5333 (2014).

162. Flory, P. J. & Rehner, J. Statistical Mechanics of Cross-Linked Polymer Networks I. Rubberlike Elasticity. *J. Chem. Phys.* **11**, 512–520 (1943).
163. Teixeira, V. G., Coutinho, F. M. B. & Gomes, A. S. Principais métodos de caracterização da porosidade de resinas à base de divinilbenzeno. *Quim. Nova* **24**, 808–818 (2001).
164. Matthias, T. *et al.* Physisorption of gases, with special reference to the evaluation of surface area and pore size distribution (IUPAC Technical Report). *Pure and Applied Chemistry* **87**, 1051 (2015).
165. Velez, J. *et al.* Simple Steps for Synthesis of Silicon Oxide Mesoporous Materials Used as Template. *J. Chil. Chem. Soc.* **58**, 1998–2000 (2013).
166. Gregg, J. S. & Sing, K. S. W. Adsorption, Surface Area and Porosity. *Acad. Press* 41 (1982). doi:10.1002/bbpc.19820861019
167. Li, J. *et al.* The supramolecular hydrogel based on hyperbranched polyglycerol and dextran as a scaffold for living cells and drug delivery. *RSC Adv.* **5**, 86730–86739 (2015).
168. Zhuo, R.-X. & Li, W. Preparation and characterization of macroporous poly(N-isopropylacrylamide) hydrogels for the controlled release of proteins. *J. Polym. Sci. Part A Polym. Chem.* **41**, 152–159 (2002).
169. Huebsch, N. & Mooney, D. J. Inspiration and application in the evolution of biomaterials. *Nature* **462**, 426–432 (2009).
170. Jaganathan, S. *et al.* Review: Radiation-induced surface modification of polymers for biomaterial application. *J. Mater. Sci.* **50**, 2007–2018 (2015).
171. Bauer, M. *et al.* In vitro hemocompatibility and cytotoxicity study of poly(2-methyl-2-oxazoline) for biomedical applications. *J. Polym. Sci. Part A Polym. Chem.* **51**, 1816–1821 (2013).
172. Wang, M. O. *et al.* Evaluation of the in vitro cytotoxicity of cross-linked biomaterials. *Biomacromolecules* **14**, 1321–1329 (2013).
173. Weber, M. *et al.* Blood-Contacting Biomaterials: In Vitro Evaluation of the Hemocompatibility. *Front. Bioeng. Biotechnol.* **6**, 99 (2018).
174. Santos, P. P., da Silva Nunes, A., Exposito de Queiroz, A. A. A. & Alencar de Queiroz, A. A. Interactions of polyglycerol dendrimers with human serum albumin: insights from fluorescence spectroscopy and computational modeling analysis. *J. Biomater. Sci. Polym. Ed.* **30**, 1575–1590 (2019).
175. Ooya, T., Ogawa, T. & Takeuchi, T. Temperature-induced recovery of a bioactive enzyme using polyglycerol dendrimers: correlation between bound water and protein interaction. *J. Biomater. Sci. Polym. Ed.* **29**, 701–715 (2018).
176. Rodrigues, M. O. *et al.* Metal organic frameworks for drug delivery and environmental remediation: A molecular docking approach. *Int. J. Quantum Chem.* **112**, 3346–3355 (2012).

177. Bourassa, P. *et al.* Locating the binding sites of anticancer tamoxifen and its metabolites 4-hydroxytamoxifen and endoxifen on bovine serum albumin. *Eur. J. Med. Chem.* **46**, 4344–4353 (2011).
178. Mandal, G., Bardhan, M. & Ganguly, T. Interaction of bovine serum albumin and albumin-gold nanoconjugates with L-aspartic acid. A spectroscopic approach. *Colloids Surfaces B Biointerfaces* **81**, 178–184 (2010).
179. Yadav, P., Bandyopadhyay, A., Chakraborty, A. & Sarkar, K. Enhancement of anticancer activity and drug delivery of chitosan-curcumin nanoparticle via molecular docking and simulation analysis. *Carbohydr. Polym.* **182**, 188–198 (2018).
180. Joshi, A. K. R., Kandlakunta, B., Kotturu, S. K. & Ghosh, S. Antiglucocorticoid potential of nutraceuticals: In silico molecular docking and in vitro assessment. *J. Food Biochem.* **42**, e12522 (2018).
181. Gupta, S. C. *et al.* Multitargeting by curcumin as revealed by molecular interaction studies. *Nat. Prod. Rep.* **28**, 1937–1955 (2011).
182. Khezri, A. *et al.* Molecular dynamic of curcumin/chitosan interaction using a computational molecular approach: Emphasis on biofilm reduction. *Int. J. Biol. Macromol.* **114**, 972–978 (2018).
183. Zhu, W. *et al.* Study of polyurethane/sulfonated dimethyl fumarate complex. *J. Appl. Polym. Sci.* **84**, 67–74 (2002).
184. Jafari, Y., Sabahi, H. & Rahaie, M. Stability and loading properties of curcumin encapsulated in *Chlorella vulgaris*. *Food Chem.* **211**, 700–706 (2016).
185. Luchtman, D. *et al.* In vivo and in vitro effects of multiple sclerosis immunomodulatory therapeutics on glutamatergic excitotoxicity. *J. Neurochem.* **136**, 971–980 (2016).
186. Vetrugno, R. *et al.* Sleep-wake and body core temperature rhythms in multiple sclerosis with fatigue. *Clin. Neurophysiol.* **118**, 228–234 (2007).
187. van de Wetering, P., Metters, A. T., Schoenmakers, R. G. & Hubbell, J. A. Poly(ethylene glycol) hydrogels formed by conjugate addition with controllable swelling, degradation, and release of pharmaceutically active proteins. *J. Control. Release* **102**, 619–627 (2005).
188. Khodaverdi, E. *et al.* Preparation and investigation of sustained drug delivery systems using an injectable, thermosensitive, in situ forming hydrogel composed of PLGA-PEG-PLGA. *AAPS PharmSciTech* **13**, 590–600 (2012).
189. Yuan, J. *et al.* Curcumin inhibits glial scar formation by suppressing astrocyte-induced inflammation and fibrosis in vitro and in vivo. *Brain Res.* **1655**, 90–103 (2017).
190. van Bochove, B. & Grijpma, D. W. Photo-crosslinked synthetic biodegradable polymer networks for biomedical applications. *J. Biomater. Sci. Polym. Ed.* **30**, 77–106 (2019).
191. Josef, E., Barat, K., Barsht, I., Zilberman, M. & Bianco-Peled, H. Composite hydrogels as a vehicle for releasing drugs with a wide range of hydrophobicities. *Acta Biomater.* **9**, 8815–8822 (2013).

192. Llabot, J., Manzo, R. & Allemandi, D. Drug release from carbomer: carbomer sodium salt matrices with potential use as mucoadhesive drug delivery system. *Int. J. Pharm.* **276**, 59–66 (2004).
193. Orakdogan, N. & Sanay, B. Dynamical modeling and experimental aspects of multi-responsive hydroxy-functional methacrylate-based gels with tunable swelling induced by multivalent ions. *Polymer (Guildf)*. **129**, 151–168 (2017).
194. Teixeira, R. S. *et al.* Effect of Cyclodextrins and pH on the permeation of tetracaine: Supramolecular assemblies and release behavior. *Int. J. Pharm.* **466**, 349–358 (2014).